

Antisynthetase syndrome: A distinct disease spectrum

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Abstract

The discovery of novel autoantibodies related to idiopathic inflammatory myopathies (collectively referred to as myositis) has not only advanced our understanding of the clinical, serological, and pathological correlation in the disease spectrum but also played a role in guiding management and prognosis. One group of the myositis-specific autoantibodies is anti-aminoacyl-tRNA synthetase (anti-ARS or anti-synthetase) which defines a syndrome with predominant interstitial lung disease, arthritis, and myositis. Autoantibodies to eight aminoacyl-tRNA synthetases have been identified with anti-Jo-1 the most common in all of idiopathic inflammatory myopathies. Disease presentation and prognosis vary depending on which anti-aminoacyl-tRNA synthetase antibody is present. In this review, we will discuss the clinical characteristics, overlap features with other autoimmune diseases, prognostic factors, and management of the antisynthetase syndrome.

Keywords

Inflammatory myositis, antisynthetase syndrome, interstitial lung disease, myositis-specific antibodies, clinicoserological spectrum

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Introduction

Idiopathic inflammatory myopathies (IIM) is a heterogeneous group of systemic autoimmune rheumatic disorders typically characterized by muscular and extra-muscular manifestations particularly skin, joint, and lung of varying severity. The presence of myositis-specific autoantibodies (MSAs) and myositis-associated autoantibodies (MAAs) has become a key feature for classification and diagnosis of IIM and is increasingly used to define clinically distinguishable IIM subsets.¹ Specific MSAs are associated with characteristic clinical phenotypes, which may assist in diagnosis, treatment, and prognostication of IIM and related complications.² Among the MSAs, autoantibodies against aminoacyl-tRNA synthetases (ARSs) were detected in 25%–35% of IIM patients.³ This review will focus on clinical characteristics, overlap features with other autoimmune diseases, prognostic factors, and management of the antisynthetase syndrome (ASSD).

ARS catalyzes the binding of a single amino acid to its specific tRNA during protein synthetase, a process that is ATP-dependent. Autoantibodies to eight ARS have been identified, namely antibodies to Jo-1 (histidyl),⁴ PL-7 (threonyl),⁵ PL-12 (alanyl),⁶ OJ (isoleucyl),⁷ EJ (glycyl),⁷ KS (asparaginy),⁸ Zo (phenylalanyl),⁹ and Ha (tyrosyl).¹

The nomenclature of anti-ARS is based on the name or initials of the index patient.¹⁰ For example, Jo-1 was named after the index patient, John P, a patient with polymyositis (PM) and interstitial lung disease (ILD), in whom anti-Jo-1 was first detected in 1980.⁴ Of the eight anti-ARS antibodies, the most common is anti-Jo-1, found in 15%–30 % of patients with IIM^{11–14} and in 60%–70% of those with ILD.¹⁰ Autoantibodies against the other ARS are less common, each less than 5% prevalence in IIM, however, collectively up to 40% of all ASSD^{14,15} (Table 1).

Although many of the anti-ARS autoantibodies have been shown to inhibit the function of their target autoantigen *in vitro*,¹⁷ the molecular pathway and the biological significance of the anti-ARS in the pathogenesis of this

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Table 1. Anti-ARS autoantibodies and prevalence.^{1,14,16}

Anti-ARS	tRNA synthetase	Prevalence in myositis (%)	Myositis ^a (%)	Arthritis ^a (%)	ILD ^a (%)
Anti-Jo-1	Histidyl	15–30	82	74	82
Anti-PL-7	Threonyl	2–5	80	50	77
Anti-PL-12	Alanyl	2–5	51	42	83
Anti-EJ	Glycyl	<2	84	53	90
Anti-OJ	Isoleucyl	<2	78	44	61
Anti-KS	Asparaginyl	<2	–	–	–
Anti-Zo	Phenylalanyl	<1	–	–	–
Anti-Ha	Tyrosyl	<1	–	–	–

ILD: interstitial lung disease.

^aAENEAS cohort of 828 patients with ASSD.¹⁶

syndrome remain elusive. Anti-nuclear antibody (ANA) has a poor sensitivity as it is only positive in half of the ASSD patients, and therefore a negative ANA should not be used to exclude this diagnosis. In contrast, ASSD patients frequently (70%–80%) have cytoplasmic staining on indirect immunofluorescence, which may serve as an important screen for ASSD patients in the right clinical setting.¹⁸

Clinical features

The cardinal clinical features of ASSD include myositis, arthritis, ILD, Raynaud's phenomenon (RP), fever, and mechanic's hands.^{15,19–21} Although the typical "triad" of arthritis, myositis, and ILD is observed in up to 90% of cases,²¹ clinically it is uncommon to see the concomitant occurrence of all three as initial presentation. Ex novo appearance of clinical features during the follow-up is a typical course of disease evolution in ASSD.^{21–25} In the American and European Network of Antisynthetase Syndrome (AENEAS) study of 225 patients with anti-Jo1-positive ASSD, only 20% patients had complete triad on presentation.²¹ Isolated arthritis, myositis, or ILD occur in up to 50% of cases. Similar findings were reported by a large cohort of Spanish patients with anti-Jo1 ASSD.²⁶ Sixty percent of the patients with incomplete ASSD developed further manifestations ex novo during the follow-up.²¹ At the end of 80-month study period, still 50% of the cohort only had one or two of the triad.²¹ Other ASSD typical clinical features, such as fever, RP, and mechanic's hands, are less frequently observed in comparison with the classic triad findings and have been reported in approximately 40% of cases.^{15,21,22} Frequency of each manifestations is summarized in Table 2. There is significant heterogeneity in clinical spectrum and time course in ASSD both within the same anti-ARS antibody and among different anti-ARS antibodies. In this section, we will summarize and highlight these unique aspects of ASSD presentation.

Table 2. Prevalence of anti-Jo-1 ASSD manifestations (%) based on AENEAS cohort data.²¹

Manifestations	Onset (%)	Follow-up (%)
Myositis	56	79
Arthritis	64	76
ILD	51	84
Complete triad	20	60
Fever	26	35
Mechanic's hands	20	30
Raynaud's phenomenon	24	37

ASSD: antisynthetase syndrome; AENEAS: American and European Network of Antisynthetase Syndrome; ILD: interstitial lung disease.

Arthritis

About 25% of ASSD presents with isolated arthritis alone, and as such, they may be misclassified as having rheumatoid arthritis (RA).^{22,25,27} In the AENEAS cohort of patients who presented with isolated arthritis, symmetric polyarticular pattern (71%) was the most common, frequently rheumatoid factor (RF)-positive (39%) and/or anti-citrullinated protein antibodies (ACPA)-positive (28%), as well as erosive changes on plain radiograph (35%),²² making RA a more likely diagnosis in the absence of other ASSD features. Indeed, 70% of these patients actually met the 1987 revised American College of Rheumatology (ACR) classification criteria for RA.²² Others found that positive ACPA in ASSD was associated with more severe and erosive arthritis,²⁸ and therefore ACPA positivity in ASSD patients may be considered as a marker of overlap with RA. In these patients, it is important to inquire respiratory and muscle symptoms even during long-term follow-up because up to 90% of patients had ex novo development of either myositis or ILD, or both, in a wide time frame, ranging from a few months to several years.²²

Not all arthritis is the same. Arthritis characteristics are heterogeneous within the syndrome and influenced by its timing of onset. Patients who have arthritis at onset of

ASSD commonly have symmetric polyarthritis (70%), whereas those with ASSD who present with *ex novo* arthritis during follow-up are much more frequently oligoarticular and asymmetrical, more likely to have ASSD accompanying findings, that is, fever, RP, and mechanic's hands, and less likely to have RA-like clinical, laboratory, and erosive radiographic features.²³

Myositis

Although ASSD is typically described as a subset of inflammatory myositis, muscle involvement is not the most frequent finding at first presentation.^{21–24} At the onset of ASSD, only 23.5% in the Pittsburgh cohort¹⁴ and 52% in the AENEAS cohort²⁹ had muscle involvement. At the end of study follow-up, still about 25% in the Pittsburgh cohort¹⁴ and 20% in the AENEAS cohort were amyopathic.²¹ In contrast, much higher rate of myositis (90%) was reported in the ASSD subset of the EuroMyositis registry, in which muscle involvement was the common reason for inclusion.³⁰ The clinical pattern of muscle involvement in ASSD is typical of IIM with subacute to chronic progressive proximal greater than distal symmetrical muscle weakness with elevated muscle enzyme and myopathic electromyography (EMG), therefore often initially diagnosed as PM. However, about 20%–25% ASSD patients have DM rashes with other typical features of myositis and therefore are clinically diagnosed as DM.

There is a considerable clinical heterogeneity among ASSD with different subtypes of anti-ARS antibodies. Jo-1 patients are much more likely to have muscle involvement than non-Jo-1 patients.¹⁴ A retrospective analysis of 166 adult Japanese patients with anti-ARS antibodies showed that myositis was well represented in those with anti-Jo-1, anti-EJ, and anti-PL-7 both at disease onset and end of follow-up period, whereas most of those with anti-PL-12, anti-KS, and anti-OJ are amyopathic throughout the entire clinical course.³¹

ILD

ILD is a hallmark feature of ASSD. The overall prevalence of ILD varies from 70% to 95% among ASSD patients depending on the cohorts.^{21,30,32} For example, in the AENEAS cohort of anti-Jo1 patients, 50% had ILD at disease onset and 84% at the end of the 80-month follow-up with three main patterns, acute/subacute, chronic, and asymptomatic, each about 1/3.²¹ The overall prevalence of ILD reached 94.4% in a Chinese ASSD cohort after a follow-up of 22 years.³² Rate of rapid progressive (RP) ILD was reported as high as 10% in this cohort.³² Similarly, in a Japanese cohort of 166 patients with ASSD, almost all eventually suffered from ILD with 7.8% RP-ILD at their first visit or during their clinical course.³¹ The higher prevalence of RP-ILD in Asian studies compared to Western

cohorts suggests perhaps a racial difference in the pulmonary manifestation of this entity. ILD has been shown to confer high mortality and morbidity in the ASSD patients.^{33,34} In the Pittsburgh ASSD cohort, 49% of mortality was attributed to ILD.¹⁴ Given high frequency and mortality associated with ILD, patients with ASSD should be screened for ILD with CT chest even if asymptomatic clinically.

There is heterogeneity among different anti-ARS antibodies in prevalence and severity of ILD. Patients with anti-PL-7 and anti-PL-12 autoantibodies more frequently have ILD³⁴ and less likely have myositis compared to anti-Jo-1 patients.²⁰ RP-ILD occurs more in the non-Jo-1 patients, particularly in those with coexisting anti-PL-7 and anti-Ro-52.³² Non-Jo-1 patients had a significantly worse unadjusted cumulative and event-free survival from diagnosis than Jo-1 patients.¹⁴ Diagnosis in non-Jo-1 patients is often delayed due to atypical or subtle presentation of connective tissue disease (CTD) features, which negatively affects their survival.¹⁴

High-resolution computed tomography (HRCT) of chest commonly reported bibasilar fibrosis, ground-glass opacities, interlobular reticulation, and traction bronchiectasis.^{35,36} Although non-specific interstitial pneumonia (NSIP) is the most common radiographic pattern followed by organizing pneumonia (OP) and usual interstitial pneumonia (UIP) in patients with ASSD associated ILD, correlation with histopathology is poor. Histologically, UIP is the most common form of ILD in anti-Jo-1,³⁷ anti-PL-7,³⁸ and anti-PL-12 cohorts.³⁹ No pathognomonic features that would allow for a definitive diagnosis of a particular ARS based on the histopathologic features alone.³⁹

Most patients with ASSD who had myositis without ILD at the onset of disease will eventually develop ILD later given the high frequency of ILD associated with ASSD.³¹ It is also important to recognize that 25% of patients with ASSD had ILD alone at disease onset and may never evolve to have muscle involvement or myositis or other characteristics.³¹ In other words, ILD alone could be the only clinical signs of ASSD.^{31,40} Except for anti-Jo-1, all the other anti-ARS antibodies are not routinely tested in the autoimmune serology panel. As a result, many patients with non-Jo-1 ASSD would otherwise be classified under idiopathic pulmonary fibrosis (IPF) or interstitial pneumonia with autoimmune features (IPAF), thus potentially entering in clinical trials addressed to other conditions or not getting appropriate treatment for ASSD. This point was illustrated by Scire et al.,²⁴ who showed 36% of patients with anti-ARS antibodies who initially met the criteria for IPAF ultimately evolved and reclassified as IIM. Many experts now suggest that patients with ILD, especially with atypical findings on CT for a classic IPF, should be tested for anti-ARS antibodies.⁴¹ Indeed, anti-ARS antibodies were found in 6%–25% of patients with IPF, many of whom had no extra-pulmonary

features.^{42–45} Whether or not skin or muscle involvement is evident, patients with anti-ARS antibodies exhibit similar ILD pattern and response to immunosuppressants.⁴¹ These patients are important to recognize, as a diagnosis of myositis-associated ILD, even for same histological subtype of UIP, is associated with a considerably better prognosis than IPF.⁴⁶ These findings further emphasize the importance of establishing clinicoserological classification criteria for ASSD.

Pulmonary artery hypertension

Pulmonary artery hypertension (PAH) was found in 11% of Jo-1 and 21% of non-Jo-1 ASSD patients in the Pittsburgh myositis cohort,¹⁴ either by right heart catheterization or echocardiography. Pulmonary hypertension is the second most common cause of death (11%) in this cohort.¹⁴ A retrospective French study noted pulmonary hypertension in one-quarter of ASSD patients by echocardiography; 8% of the cohort was confirmed severe PAH by right heart catheterization.⁴⁷ This was likely an underestimate as only selected few underwent right heart catheterization. Presence of pulmonary hypertension was significantly associated with a lower survival rate, with a 3-year survival rate of 58%.⁴⁷ Pulmonary hypertension is under-recognized in patients with ASSD. We recommend doing screening echocardiogram at baseline even for asymptomatic individuals with ASSD and subsequently on follow-up for any evidence of exercise-induced desaturation or dyspnea.⁴⁸

Other accompanying features

Mechanic's hand represents the typical cutaneous manifestation of ASSD.⁴⁹ The novel finding of Hiker's feet is the equivalent of mechanic's hands, newly recognized in the recent years.^{50,51} These are characterized by hyperkeratotic scaly fissured plaques on the fingers/palms or soles, generally more evident on the lateral aspect of index fingers.^{52,53} Mechanic's hand is suggestive of but not specific to ASSD and can be found in IIM associated with other autoantibodies.^{31,54} Presence of mechanic's hand in ASSD is associated with a higher rate of ILD in some studies.^{53,55,56}

RP is another non-specific symptom associated with ASSD although the exact mechanism of vasculopathy in ASSD is unclear. RP is more prevalent in ASSD associated with PL-7 and PL-12 in comparison with Jo-1,⁵⁷ and rarely, cases of recurrent digital ulcers have been reported.⁵⁸ Nailfold video capillaroscopy (NVC) abnormality can be seen in more than 60% of patients with ASSD.⁵⁹ The presence of a systemic sclerosis (SSc)-like pattern is common in ASSD especially with anti-Jo-1 antibody but is independent from the occurrence of RP,⁵⁹ suggesting nailfold exam perhaps should be a routine part of ASSD evaluation even without RP symptoms.

Other than the classic features mentioned above, calcinosis, panniculitis, coronary artery dilatation, eosinophilic pleural effusions, and myopericarditis have also been reported in small case series.^{60–69}

Overlap with other CTDs

Overlap between IIM and other CTD is common. The immunological profile is generally characterized by the presence of autoantibodies which are not specific to but frequently associated with myositis. Anti-PM/Scl, anti-Ku as well as anti-U1 RNP and anti-SSA/Ro represent the most frequent MAAs. The analysis of seven large cohorts in the literature showed that 6.5%–36.7% of the patients with IIM presented overlap syndrome,^{70–75} most commonly SSc, approximately 30%–50%^{70,76,77} also known as scleromyositis. In SSc cohort, about 6%–8% have SSc-myositis overlap.^{78,79}

However, ASSD overlap with SSc is not commonly reported and has been shown only in a few case reports in PL-7, PL-12, and Jo-1.^{70,80} In a Dutch SSc cohort, only 2 of the 422 patients with SSc patients had Jo-1 antibody.⁷⁸ Although the pathogenesis of SSc and ASSD seems to be quite different, vasculopathy is a typical manifestation shared by both diseases. De novo development of sclerodactyly, RP, and finger-tip ulcerations with positive RNP antibody was reported in a case of established anti-PL12 ASSD. Scleroderma-like renal crisis with biopsy evidence of thrombotic microangiopathy was described in one case of new diagnosis of anti-PL-7 ASSD who had negative usual SSc antibodies (ENA, RNA polymerase III, Scl, and centromere).⁸¹

Histopathological features of the muscle biopsy

Clinico-sero-pathological correlation is increasingly recognized. ASSD is a distinctive pathological subset among IIM. The hallmark of ASSD in muscle pathology is perifascicular necrosis (necrosis and regenerating fibers located at the periphery of the muscle fascicle),⁸² although diffusely distributed necrotic and regenerating fibers not restricted to the perifascicular area have also been described in a large cohort of 50 ASSD patients.⁸³ Very frequently, there is increased expression of major histocompatibility complex (MHC) class I and class II in the cytoplasm and sarcolemma fibers and sarcolemmal membrane attack complex deposition, predominantly in the perifascicular region.^{83–86} Other findings also include perimysial connective tissue fragmentation and alkaline phosphatase overexpression in perimysium.^{83,87,88} ASSD muscle biopsy can be sometimes be misclassified as dermatomyositis (DM), which is characterized by the presence of perifascicular inflammation, myofiber atrophy, MHC upregulation,^{89,90} and immune-mediated necrotizing myopathy (IMNM), which is characterized by diffuse

muscle necrosis with little to no endomysial lymphocytic infiltration.⁸⁹

Anti-Ro 52 in ASSD

Of the MAAs, anti-Ro 52 antibody draws much attention with regard to its importance in both prognosis and therapeutic implication. The cytoplasmic Ro/SSA antigen has two polypeptide components: Ro 52 and Ro 60, autoantibodies to both of which are commonly seen in CTD. However, Ro 52 is biochemically and immunologically distinct from Ro 60 and is considered more immunogenic.⁹¹ Anti-Ro 52 is present in about 30% of IIM⁹² and has a strong association with anti-ARS antibodies in comparison with other MSAs or MSA negative DM.^{1,93,94} It was found in 40%–72% of patients with anti-Jo-1 ASSD and up to 70% in those with non-Jo-1 ASSD depending on the population studied, and there was no associated concomitant Sjögren syndrome clinically.^{11,93–97} Some reported that patients with anti-PL7, anti-PL12, and anti-EJ were much more likely to have anti-Ro 52 than those with anti-Jo-1.³²

Presence of anti-Ro 52 in patients with ASSD has been related to more severe ILD, relapses, and refractory disease^{97–99}; however, correlation of this autoantibody with degree of myositis, arthritis, or skin manifestations is inconsistent between studies.^{95,100} Acute onset respiratory failure and development of lung fibrosis was more frequently observed in ASSD with anti-Ro 52 than those without anti-Ro 52, particularly in anti-PL-7 positive patients,³² with high anti-Ro 52 concentrations showing the highest risk.¹⁰⁰ Similar effect of anti-Ro 52 on ILD has also been reported in association with other MSAs particularly anti-MDA5.¹⁰¹

At least in anti-Jo-1 ASSD, presence of high-concentration anti-Ro 52 also confers significance in therapeutic response. Bauhammer et al. showed patients with high anti-Ro 52 titer responded poorly to conventional immunosuppressants including cyclophosphamide (CYC) and cyclosporin but all responded well to rituximab (RTX). In contrast, those with low-titer anti-Ro 52 did well on conventional immunosuppressants alone.^{95,100}

The mechanism as to why concomitant anti-Ro 52 with anti-ARS leads to more severe clinical phenotype remains elusive. Ro 52 is also known as TRIM21 (tripartite motif proteins). It is an interferon (IFN)-inducible E3 ligase that mediates the ubiquitination and proteasomal degradation of various IFN regulatory factor transcription factors, leading to a downregulation of Type-I IFN.¹⁰² It is postulated that antibody to Ro 52 suppresses the degradation and therefore excessive production of proinflammatory cytokines.¹⁰²

Malignancy in ASSD

Previously, a negative association between ILD, especially in ASSD patients, and cancer was accepted. In a

large retrospective analysis of 233 patients with ASSD, the frequency of cancer was only 1.7% within 3 years from ASSD diagnosis, in which the authors assert was not much different than the general population in France.²⁰ However, other case studies and review demonstrated a significant increased risk of cancer in these patients, ranging from 4% to 13% of ASSD patients.^{26,32,103,104} Additional cases and small series have also been published.^{19,20,105–113} Most frequently identified cancer types are lung and colon, and less frequently breast and ovarian.¹⁰⁵ Only one study reported association of positive anti-Ro 52 with higher prevalence of malignancy.⁹⁵ Malignancy is associated with poor prognosis in ASSD.³² These emerging data support that even though overall cancer risk may be lower than other autoantibody subsets, still a diligent malignancy screen for ASSD patients is warranted.

Outcomes in ASSD

Some negative prognostic factors have been recently identified in ASSD. The presence of anti-Ro 52, RP-ILD, PH, and malignancy have been identified to associate with poor prognosis.³² Others showed male sex, reduced DLCO (diffusing capacity of lung for carbon monoxide) at presentation, and elevated serum ferritin correlated with reduced survival.^{32,36,114} Black American ethnicity is also an independent prognostic factor associated with increased lung involvement severity.¹¹⁵

Reported survival rate of ASSD patients are fairly similar between various studies.^{14,26,32,36} The Pittsburgh myositis registry of 202 ASSD patients followed over 25 years (1985–2009) showed overall 33% mortality and 6% required lung transplantation. The 5- and 10-year unadjusted cumulative survival rates from diagnosis were 84% and 61%, respectively.¹⁴ Analysis from a large Chinese cohort of 128 ASSD patients showed cumulative 10-year survival rate of 76.8%³² and similarly, the Spanish cohort of 148 anti-Jo-1 ASSD patients had 25% mortality after a median follow-up of 6 years.²⁶

Among different anti-ARS antibodies, anti-non-Jo-1 ASSD have worse clinical outcome from anti-Jo-1 patients.^{14,116,117} The anti-Jo-1 patients had a significantly better 5- and 10-year unadjusted cumulative survival from diagnosis (90% and 70%, respectively) as compared to non-Jo-1 patients (75% and 47%, respectively).¹⁴ Several variables affect the survival difference between anti-Jo-1 and non-Jo-1 patients. A major factor is the delay in diagnosis as anti-non-Jo-1 patients often present atypically with more frequent ILD³⁴ and non-specific presentation, and less frequently myositis or arthritis.²⁰ Second, anti-non-Jo-1 antibodies are not routinely tested or available, and when tested, the reliability of commercial testing is highly variable. Finally, anti-non-Jo-1 ASSD, particularly anti-PL7, is associated with more frequent RP-ILD which confers poor prognosis.³²

Table 3. Recommended measures and investigations at diagnosis and each follow-up.

Domains	Suggested measures and investigations	
	At diagnosis	At each follow-up
General	History and examination to identify involved systems	Interval history and repeat examination to assess treatment responses
Muscle	MMT8 and consider: MRI pelvic/thighs and shoulder girdle muscles EMG Muscle biopsy	MMT8
Joint	TJC, SJC	
Lung	CXR, HRCT, PFT, 6MWT, echocardiogram for PAH if isolated decrease in DLCO, referral to respiratory for co-management if ILD	HRCT, PFT, and 6MWT frequency determined by progression of disease, often every 3–12 months directed by respiratory
Skin	General assessment and consider documentation with CDASI Referral to dermatologists	
Raynaud's phenomenon	Conservative and pharmacological management review	
Bloodwork	CBC, creatinine, CK, LDH, aldolase, AST, ALT, ANA, ENA, RF, ACPA, MSAs and MAAs	CBC, creatinine, CK, LDH, aldolase, AST, ALT
Overall disease activity and damage assessment	Consider MDAAT, MDI	
Patient-reported outcomes	HAQ, SF-36	

TJC: tender joint count; SJC: swollen joint count; PAH: pulmonary artery hypertension; MMT8: manual muscle testing 8 (https://www.niehs.nih.gov/research/resources/assets/docs/mmt8_grading_and_testing_procedures_for_the_abbreviated_8_muscle_groups_508.pdf); MRI: magnetic resonance imaging; EMG: electromyography; CXR: chest X-ray; HRCT: high-resolution computed tomography; PFT: pulmonary function test; 6MWT: 6-minute walk test; DLCO: diffusing capacity of lung for carbon monoxide; ILD: interstitial lung disease; CDASI: cutaneous dermatomyositis disease area and severity index; CBC: complete blood count; CK: creatine kinase; LDH: lactate dehydrogenase; AST: aspartate aminotransferase; ALT: alanine transaminase; ANA: antinuclear antibody; ENA: extractable nuclear antigens; RF: rheumatoid factor; ACPA: anti-citrullinated protein antibody; MSA: myositis-specific antibody; MAA: myositis-associated antibody; MDAAT: Myositis Disease Activity Assessment Tool; MDI: myositis damage index; HAQ: Health Assessment Questionnaire; SF-36: Short-Form 36 Health Survey Questionnaire.

Treatment in ASSD: focus on ILD

Managing ASSD represents a challenge to the clinicians as patients can have multi-system involvement, ILD being the most life-threatening. Muscle, skin, joint, and lung disease activity in ASSD may have different clinical course and response to therapy and often require multidisciplinary approach from rheumatologists, dermatologists, and pulmonologists. In Table 3, we recommend to thoroughly review patients and arrange investigations by organ systems at first visit and during each follow-up.

ILD is a major cause of morbidity and mortality in myositis requiring combinations of glucocorticoids, immunosuppressive drugs, and agents that modulate T-cell function and deplete B-cells. Unlike the antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis literature, there is little evidence in choice of drugs or duration of therapy in “induction” and “maintenance.” There are very few large treatment trial results available to guide myositis treatment and none for myositis-associated ILD. We will focus the discussion on myositis-associated ILD treatment with emphasis on ASSD in this section. A proposed treatment algorithm is illustrated in Figure 1.

Corticosteroids have long been first-line in the treatment of IIM but monotherapy with corticosteroid alone is rarely sufficient in ASSD, as ILD often recurs with corticosteroid

tapering. Additional immunosuppressive agents are added for refractory skin, muscle, and/or lung disease as corticosteroid-sparing agents. There is no consensus or guidelines for additional immunosuppressive drugs, but frequently used agents include azathioprine, mycophenolate mofetil (MMF), tacrolimus, RTX, and CYC, which are summarized in Table 4. Methotrexate is very effective for ASSD with arthritis and/or myositis, but clinicians generally avoid methotrexate if ILD is also a predominant symptom, primarily due to potential difficulty in ascertaining the cause of worsening ILD to disease versus methotrexate pneumonitis. There is no evidence of increased methotrexate pneumonitis in non-RA diseases.¹¹⁹ Intravenous immunoglobulin (IVIG) is beneficial when there is esophageal involvement, refractory rash, pregnancy, active infection, and calcifications but evidence of its use in myositis-associated ILD is limited to case reports only.^{120,121}

Azathioprine treatment led to modest improvement in lung function in small retrospective case series or cohorts.^{122,123,147} Significant improvements in forced vital capacity (FVC) and DLCO were demonstrated in a large retrospective study of 125 patients with CTD-associated ILD (including 32 myositis patients) who were treated with MMF.¹²⁴ Benefit of CYC (IV or oral) in myositis-associated ILD has primarily been reported in retrospective case reports and case series.^{134–136} Clinically significant

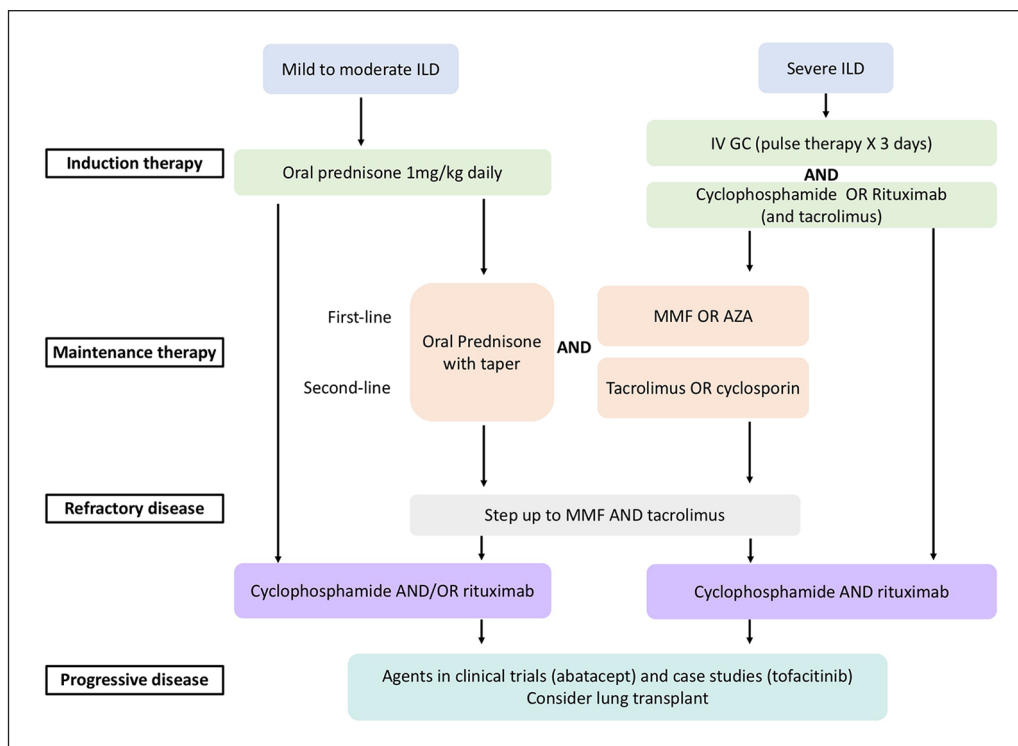


Figure 1. A proposed treatment algorithm for myositis-associated ILD (adapted from Oddis and Aggarwal¹¹⁸).

Table 4. Steroid-sparing agents for myositis-associated ILD.

Drug	Dose	Mechanism of action	Indication	Evidence
Established therapies				
Azathioprine	2–3 mg/kg orally	Convert into 6-mercaptopurine which halt DNA replication and block purine synthesis	Most commonly used steroid-sparing agent	Retrospective case series ^{122,123}
Mycophenolate mofetil	1–1.5 g in divided dose orally	Convert into mycophenolic acid to inhibit guanosine nucleotide synthetase which in turn inhibits T- and B-cell proliferation	Increasing evidence in myositis ILD and refractory cutaneous disease	Retrospective case series and cohort ^{124,125–128}
Tacrolimus	1 mg orally twice a day and titrate to reach trough level of 5–10 ng/mL	Binds to FKBP12, leading to inhibition of T cell activation	Increasing evidence in myositis ILD Add on therapy for rapid progressive ILD	Retrospective studies ^{129–133}
Cyclophosphamide	Oral 1.5–2 mg/kg/day or IV 500–1000 mg/m ² monthly, or IV 10–15 mg/kg every 3–4 weeks	An alkylating agent that interferes with the growth of cells	Refractory or rapidly progressive ILD	Retrospective case reports and series ^{134–136} and one open-label study ¹³⁷
Rituximab	Two 1 g doses 15 days apart, but maintenance dose and interval may vary	Depletes CD20 + B cells	Refractory or rapidly progressive ILD	Retrospective case reports and one open-label trial ^{100,138–140}
Potential therapies				
Tofacitinib	5 mg twice a day orally	Inhibits JAK enzymes and downstream STATs	Add on therapy in refractory cutaneous DM and myositis-associated ILD	Only case reports and case series ^{141–145}
Abatacept	125 mg subcutaneous weekly	Inhibits T-cell activation by binding to CD80 and CD86	Currently in a phase II pilot study for ASSD ILD	Trial underway ¹⁴⁶

ILD: interstitial lung disease; JAK: Janus kinase; DM: diabetes mellitus; ASSD: antisynthetase syndrome.

improvement was seen in dyspnea scale, oxygen requirement, FVC, and HRCT of the lung parenchyma.¹³⁶ In a prospective open-label study of 20 patients with myositis-associated ILD which included 10 RP-ILD, all 10 patients received IV CYC 10–12 mg/kg every 3 weeks for total 6–9 doses and resulted in stabilization, functional improvement, as well as increase in VC, FVC, and DLCO measures, with a median follow-up of 35 months.¹³⁷ Based on these evidences, we recommend IV CYC for severe and progressive myositis-associated ILD, in addition to the pulse glucocorticoid.

There is growing evidence of T-cell inhibition in the management of myositis-associated ILD. Cyclosporine and tacrolimus are the two available calcineurin inhibitors. For those with myositis-associated ILD, in whom cyclosporine was ineffective, small case series showed that tacrolimus treatment may improve lung disease.^{129–131} In vitro tacrolimus is 50-fold more potent than cyclosporine,¹⁴⁸ therefore tacrolimus serum trough levels should be monitored to ensure therapeutic level and to limit toxicity. In a retrospective review of 13 ASSD patients with ILD (12 anti-Jo-1 and 1 anti-PL-12) who were treated with tacrolimus for an average of 51 months, a significant improvement was observed in all pulmonary parameters measured as well as decline in creatine kinase (CK) and increase in muscle strength.¹³¹ Calcineurin inhibitors have been shown to be effective in improving pulmonary function (PFT—pulmonary function test) and HRCT of chest¹³² and should be used early¹⁴⁹ in progressive myositis-associated ILD. Asian studies have shown that addition of calcineurin inhibitor to conventional therapy led to longer event-free survival than treatment with conventional therapy alone.^{133,150,151}

Increasing number of retrospective studies have shown benefit and safety of RTX in ILD associated with ASSD with reported objective improvement in PFT, ground-glass opacities and stability, or fibrosis on HRCT of chest.^{100,138,139,152–160} Those who had acute onset of ILD or had RTX within 12 months of disease onset had the best outcome.¹³⁹ In an open-label study of 10 patients with ASSD and refractory ILD, treatment with RTX resulted in a substantial steroid-sparing effect; clinically significant increase in FVC and/or DLCO was observed in 5 out of the 10 patients, stabilization in 4, and worsening in 1.¹³⁸ A retrospective study of 25 ASSD-ILD subjects treated with RTX from 2 US academic referral centers showed significant steroid-sparing effect, good tolerance, and improvement in CT imaging and/or PFT in most patients at 1- and 3-year follow-up.¹⁵⁹ In another retrospective study of 17 anti-Jo-1-positive ASSD patients who received RTX, 16 showed a more rapid and marked response than the others who were treated with conventional immunosuppressants (including CYC, cyclosporine, azathioprine, methotrexate, and leflunomide). This study also confirmed the effectiveness of RTX in those with refractory ILD, particularly in cases of concomitant high titers of anti-Ro 52.¹⁰⁰ Although

not specific for ILD outcome, presence of ARS antibodies was predictive of better outcome in the post hoc analysis of the RTX in myositis trial.¹⁶¹

In all of these studies, RTX is usually administered as two 1-g doses 2 weeks apart. Some supports to repeat RTX dosing over a single RTX cycle¹⁵⁹ but the subsequent dosing interval might vary, and no regimen has been standardized for the treatment of myositis with or without ILD. The RECITAL trial which is currently recruiting is the first randomized control trial to study the efficacy of RTX versus CYC as first-line treatment in CTD-associated ILD.¹⁶²

Tofacitinib has been shown beneficial in refractory cutaneous manifestation of DM.^{141,142} Its benefit as an additional therapy to CYC or RTX in refractory ILD was also demonstrated in recent case reports and case series.^{143–145} Abatacept is currently being assessed in a randomized controlled phase II trial to evaluate its efficacy, safety, and tolerability in ASSD associated ILD.

Treatment of other manifestations including myositis, arthritis, and cutaneous lesions parallels that of ILD summarized in Table 4. For refractory myositis or DM rash, IVIG can be effective.¹⁶³ For patients presented with predominant arthritis or cutaneous disease with no or minimal ILD, methotrexate or leflunomide can also be considered.

Summary

ASSD, although characterized by the classic triad of myositis, arthritis, and ILD, often presents with incomplete forms with only one or two of the manifestations in the presence of anti-ARS antibodies. Myositis in ASSD is clinically typical of PM (sometimes DM if rashes are present), but histopathologically distinct. Arthritis can be presenting symptoms and is often misdiagnosed as RA. ILD is the most common and often the presenting clinical feature in ASSD and is highly associated with higher mortality. Given that the anti-ARS antibodies are found in 6%–25% of idiopathic ILD, we consider testing anti-ARS in any ILD patients with or without other clinical features, as the presence of these antibodies confers different treatment and prognosis. PAH is common in ASSD; together with ILD, PAH confers a significant pulmonary mortality. We recommend doing screening echocardiogram at baseline for all ASSD patients and on follow-up for any evidence of exercise-induced desaturation or decline in DLCO on routine PFTs. For treatment of ASSD, conventional therapies include glucocorticoids usually in combination with one or multiple immunosuppressive drugs (DMARDs—disease-modifying antirheumatic drugs) like azathioprine, methotrexate, and MMF. Given the high prevalence of ILD, immunosuppressive agents that modulate T-cell function such as tacrolimus and deplete B-cells (RTX) are often required and helpful.

There is growing recognition that clinicoserological classification criteria for ASSD are needed. The new 2017 ACR/

EULAR (European League Against Rheumatism) criteria do not capture the ASSD especially because the criteria weigh heavily on cutaneous and muscular findings which may be absent or evolve later in ASSD patients.¹⁶⁴ Presence of ILD or non-Jo-1 antibodies is not a part of the criteria. To overcome the shortcomings of 2017 EULAR/ACR myositis criteria and to promote clinical classification and early recognition of ASSD, currently there is a large multi-center international ACR/EULAR funded project for development and validation of ASSD classification criteria.

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References

- Ghirardello A, Bassi N, Palma L, et al. Autoantibodies in polymyositis and dermatomyositis. *Curr Rheumatol Rep* 2013; 15: 335.
- Targoff IN. Idiopathic inflammatory myopathy: autoantibody update. *Curr Rheumatol Rep* 2002; 4(5): 434–441.
- Koenig M, Fritzler MJ, Targoff IN, et al. Heterogeneity of autoantibodies in 100 patients with autoimmune myositis: insights into clinical features and outcomes. *Arthritis Res Ther* 2007; 9(4): R78.
- Nishikai M and Reichlin M. Heterogeneity of precipitating antibodies in polymyositis and dermatomyositis. Characterization of the Jo-1 antibody system. *Arthritis Rheum* 1980; 23: 881–888.
- Mathews MB, Reichlin M, Hughes GR, et al. Anti-threonyl-tRNA synthetase, a second myositis-related autoantibody. *J Exp Med* 1984; 160(2): 420–434.
- Bunn CC, Bernstein RM and Mathews MB. Autoantibodies against alanyl-tRNA synthetase and tRNAAla coexist and are associated with myositis. *J Exp Med* 1986; 163(5): 1281–1291.
- Targoff IN. Autoantibodies to aminoacyl-transfer RNA synthetases for isoleucine and glycine. Two additional synthetases are antigenic in myositis. *J Immunol* 1990; 144(5): 1737–1743.
- Hirakata M, Suwa A, Nagai S, et al. Anti-KS: identification of autoantibodies to asparaginyl-transfer RNA synthetase associated with interstitial lung disease. *J Immunol* 1999; 162(4): 2315–2320.
- Betteridge Z, Gunawardena H, North J, et al. Antisynthetase syndrome: a new autoantibody to phenylalanyl transfer RNA synthetase (anti-Zo) associated with polymyositis and interstitial pneumonia. *Rheumatology (Oxford)* 2007; 46(6): 1005–1008.
- Mahler M and Fritzler MJ. Epitope specificity and significance in systemic autoimmune diseases. *Ann N Y Acad Sci* 2010; 1183: 267–287.
- Brouwer R, Hengstman GJ, Vree Egberts W, et al. Autoantibody profiles in the sera of European patients with myositis. *Ann Rheum Dis* 2001; 60(2): 116–123.
- Vazquez-Abad D and Rothfield NF. Sensitivity and specificity of anti-Jo-1 antibodies in autoimmune diseases with myositis. *Arthritis Rheum* 1996; 39(2): 292–296.
- Arnett FC, Targoff IN, Mimori T, et al. Interrelationship of major histocompatibility complex class II alleles and autoantibodies in four ethnic groups with various forms of myositis. *Arthritis Rheum* 1996; 39(9): 1507–1518.
- Aggarwal R, Cassidy E, Fertig N, et al. Patients with non-Jo-1 anti-tRNA-synthetase autoantibodies have worse survival than Jo-1 positive patients. *Ann Rheum Dis* 2014; 73(1): 227–232.
- Chatterjee S, Prayson R and Farver C. Antisynthetase syndrome: not just an inflammatory myopathy. *Cleve Clin J Med* 2013; 80: 655–666.
- Cavagna L, Trallero-Araguas E, Meloni F, et al. Influence of antisynthetase antibodies specificities on antisynthetase syndrome clinical spectrum time course. *J Clin Med* 2019; 8(11): E2013.
- Targoff IN, Trieu EP and Miller FW. Reaction of anti-OJ autoantibodies with components of the multi-enzyme complex of aminoacyl-tRNA synthetases in addition to isoleucyl-tRNA synthetase. *J Clin Invest* 1993; 91(6): 2556–2564.
- Aggarwal R, Dhillon N, Fertig N, et al. A negative antinuclear antibody does not indicate autoantibody negativity in myositis: role of anticypoplasmic antibody as a screening test for antisynthetase syndrome. *J Rheumatol* 2017; 44: 223–229.
- Dugar M, Cox S, Limaye V, et al. Clinical heterogeneity and prognostic features of South Australian patients with anti-synthetase autoantibodies. *Intern Med J* 2011; 41(9): 674–679.
- Hervier B, Devilliers H, Stanciu R, et al. Hierarchical cluster and survival analyses of antisynthetase syndrome: phenotype and outcome are correlated with anti-tRNA synthetase antibody specificity. *Autoimmun Rev* 2012; 12(2): 210–217.
- Cavagna L, Nuno L, Scire CA, et al. Clinical spectrum time course in anti Jo-1 positive antisynthetase syndrome: results from an international retrospective multicenter study. *Medicine (Baltimore)* 2015; 94(32): e1144.
- Cavagna L, Nuno L, Scire CA, et al. Serum Jo-1 autoantibody and isolated arthritis in the antisynthetase syndrome: review of the literature and report of the experience of AENEAS collaborative group. *Clin Rev Allergy Immunol* 2017; 52(1): 71–80.
- Gonzalez-Gay MA, Montecucco C, Selva-O'Callaghan A, et al. Timing of onset affects arthritis presentation pattern in antisynthetase syndrome. *Clin Exp Rheumatol* 2018; 36(1): 44–49.
- Scire CA, Gonzalez-Gay MA, Selva-O'Callaghan A, et al. Clinical spectrum time course of interstitial pneumonia with autoimmune features in patients positive for antisynthetase antibodies. *Respir Med* 2017; 132: 265–266.

25. Bartoloni E, Gonzalez-Gay MA, Scire C, et al. Clinical follow-up predictors of disease pattern change in anti-Jo1 positive anti-synthetase syndrome: results from a multicenter, international and retrospective study. *Autoimmun Rev* 2017; 16(3): 253–257.
26. Trallero-Araguas E, Grau-Junyent JM, Labirua-Iturburu A, et al. Clinical manifestations and long-term outcome of anti-Jo1 antisynthetase patients in a large cohort of Spanish patients from the GEAS-IIM group. *Semin Arthritis Rheum* 2016; 46(2): 225–231.
27. Lefevre G, Meyer A, Launay D, et al. Seronegative polyarthritis revealing antisynthetase syndrome: a multicentre study of 40 patients. *Rheumatology (Oxford)* 2015; 54(5): 927–932.
28. Meyer A, Lefevre G, Bierry G, et al. In antisynthetase syndrome, ACPA are associated with severe and erosive arthritis: an overlapping rheumatoid arthritis and antisynthetase syndrome. *Medicine (Baltimore)* 2015; 94(20): e523.
29. Cavagna L, Castaneda S, Scire C, et al. Antisynthetase syndrome or what else? Different perspectives indicate the need for new classification criteria. *Ann Rheum Dis* 2018; 77: e50.
30. Lilleker JB, Vencovsky J, Wang G, et al. The EuroMyositis registry: an international collaborative tool to facilitate myositis research. *Ann Rheum Dis* 2018; 77(1): 30–39.
31. Hamaguchi Y, Fujimoto M, Matsushita T, et al. Common and distinct clinical features in adult patients with anti-aminoacyl-tRNA synthetase antibodies: heterogeneity within the syndrome. *PLoS ONE* 2013; 8(4): e60442.
32. Shi J, Li S, Yang H, et al. Clinical profiles and prognosis of patients with distinct antisynthetase autoantibodies. *J Rheumatol* 2017; 44(7): 1051–1057.
33. Johnson C, Connors GR, Oaks J, et al. Clinical and pathologic differences in interstitial lung disease based on antisynthetase antibody type. *Respir Med* 2014; 108(10): 1542–1548.
34. Marie I, Josse S, Decaux O, et al. Comparison of long-term outcome between anti-Jo1- and anti-PL7/PL12 positive patients with antisynthetase syndrome. *Autoimmun Rev* 2012; 11(10): 739–745.
35. Waseda Y, Johkoh T, Egashira R, et al. Antisynthetase syndrome: pulmonary computed tomography findings of adult patients with antibodies to aminoacyl-tRNA synthetases. *Eur J Radiol* 2016; 85(8): 1421–1426.
36. Zamora AC, Hoskote SS, Abascal-Bolado B, et al. Clinical features and outcomes of interstitial lung disease in anti-Jo-1 positive antisynthetase syndrome. *Respir Med* 2016; 118: 39–45.
37. Richards TJ, Eggebeen A, Gibson K, et al. Characterization and peripheral blood biomarker assessment of anti-Jo-1 antibody-positive interstitial lung disease. *Arthritis Rheum* 2009; 60(7): 2183–2192.
38. Yousem SA, Schneider F, Bi D, et al. The pulmonary histopathologic manifestations of the anti-PL7/antithreonyl transfer RNA synthetase syndrome. *Hum Pathol* 2014; 45(6): 1199–1204.
39. Schneider F, Yousem SA, Oddis CV, et al. Pulmonary pathologic manifestations of anti-alanyl-tRNA synthetase (anti-PL-12)-related inflammatory myopathy. *Arch Pathol Lab Med* 2018; 142(2): 191–197.
40. Ghysen K and Leys M. A 64-year-old woman with interstitial lung disease and positive antibodies against aminoacyl-transfer RNA synthetases in the absence of myositis: presentation of an anti-PL-12 positive antisynthetase syndrome. *Acta Clin Belg* 2018; 73(5): 389–392.
41. Takato H, Waseda Y, Watanabe S, et al. Pulmonary manifestations of anti-ARS antibody positive interstitial pneumonia—with or without PM/DM. *Respir Med* 2013; 107(1): 128–133.
42. Matsushita T, Hasegawa M, Fujimoto M, et al. Clinical evaluation of anti-aminoacyl tRNA synthetase antibodies in Japanese patients with dermatomyositis. *J Rheumatol* 2007; 34(5): 1012–1018.
43. Nakashima R, Imura Y, Hosono Y, et al. The multicenter study of a new assay for simultaneous detection of multiple anti-aminoacyl-tRNA synthetases in myositis and interstitial pneumonia. *PLoS ONE* 2014; 9(1): e85062.
44. Watanabe K, Handa T, Tanizawa K, et al. Detection of antisynthetase syndrome in patients with idiopathic interstitial pneumonias. *Respir Med* 2011; 105(8): 1238–1247.
45. Yura H, Sakamoto N, Satoh M, et al. Clinical characteristics of patients with anti-aminoacyl-tRNA synthetase antibody positive idiopathic interstitial pneumonia. *Respir Med* 2017; 132: 189–194.
46. Aggarwal R, McBurney C, Schneider F, et al. Myositis-associated usual interstitial pneumonia has a better survival than idiopathic pulmonary fibrosis. *Rheumatology (Oxford)* 2017; 56(3): 384–389.
47. Hervier B, Meyer A, Dieval C, et al. Pulmonary hypertension in antisynthetase syndrome: prevalence, aetiology and survival. *Eur Respir J* 2013; 42(5): 1271–1282.
48. Chatterjee S and Farver C. Severe pulmonary hypertension in anti-Jo-1 syndrome. *Arthritis Care Res (Hoboken)* 2010; 62(3): 425–429.
49. Gusdorf L, Morrucci C, Goetz J, et al. Mechanics hands in patients with antisynthetase syndrome: 25 cases. *Ann Dermatol Venereol* 2019; 146: 19–25.
50. Cox JT, Gullotti DM, Mecoli CA, et al. “Hiker’s feet”: a novel cutaneous finding in the inflammatory myopathies. *Clin Rheumatol* 2017; 36(7): 1683–1686.
51. Wernham M and Montague SJ. Mechanic’s hands and hiker’s feet in antisynthetase syndrome. *CMAJ* 2017; 189(44): E1365.
52. Katzap E, Barilla-LaBarca ML and Marder G. Antisynthetase syndrome. *Curr Rheumatol Rep* 2011; 13: 175–181.
53. Mii S, Kobayashi R, Nakano T, et al. A histopathologic study of mechanic’s hands associated with dermatomyositis: a report of five cases. *Int J Dermatol* 2009; 48(11): 1177–1182.
54. Sohara E, Saraya T, Sato S, et al. Mechanic’s hands revisited: is this sign still useful for diagnosis in patients with lung involvement of collagen vascular diseases? *BMC Res Notes* 2014; 7: 303.
55. Patel HC and Lauder NN. The antisynthetase syndrome. *Am J Med* 2011; 124: e3–e4.
56. Bachmeyer C, Tillie-Leblond I, Lacert A, et al. “Mechanic’s hands”: a misleading cutaneous sign of the antisynthetase syndrome. *Br J Dermatol* 2007; 156(1): 192–194.

57. Mercer LK, Moore TL, Chinoy H, et al. Quantitative nailfold video capillaroscopy in patients with idiopathic inflammatory myopathy. *Rheumatology (Oxford)* 2010; 49(9): 1699–1705.
58. Miyagawa F, Azukizawa H, Mimori T, et al. Anti-PL-12 antibody-positive antisynthetase syndrome with recurrent digital ulcers. *J Dermatol* 2018; 46: e143–e145.
59. Sebastiani M, Triantafyllias K, Manfredi A, et al. Nailfold capillaroscopy characteristics of antisynthetase syndrome and possible clinical associations: results of a multicenter international study. *J Rheumatol* 2019; 46: 279–284.
60. Asi K, Gourishankar A and Kamdar A. Coronary artery dilation associated with anti-synthetase syndrome in an adolescent. *Pediatr Rheumatol Online J* 2019; 17(1): 3.
61. Hayashi K, Machida Y, Katayama Y, et al. A case of antisynthetase syndrome with anti-EJ antibody complicated by pericarditis. *Rins Shinkeig* 2019; 59: 21–26.
62. Labirua-Iturburu A, Selva-O’Callaghan A, Vincze M, et al. Anti-PL-7 (anti-threonyl-tRNA synthetase) antisynthetase syndrome: clinical manifestations in a series of patients from a European multicenter study (EUMYONET) and review of the literature. *Medicine (Baltimore)* 2012; 91(4): 206–211.
63. Mehndiratta P, Mehta S, Manjila SV, et al. Isolated necrotizing myopathy associated with ANTI-PL12 antibody. *Muscle Nerve* 2012; 46(2): 282–286.
64. Meudec L, Jelin G, Forien M, et al. Antisynthetase syndrome and cardiac involvement: a rare association. *Joint Bone Spine*. Epub ahead of print 7 October 2018. DOI: 10.1016/j.jbspin.2018.09.019.
65. Rungcharunthanakul A and Sampattavanich N. Polymyositis with antisynthetase syndrome presented with lobular panniculitis: a case report. *Clin Case Rep* 2019; 7(1): 128–130.
66. Dieval C, Deligny C, Meyer A, et al. Myocarditis in patients with antisynthetase syndrome: prevalence, presentation, and outcomes. *Medicine (Baltimore)* 2015; 94(26): e798.
67. Saito G, Kono M, Tsutsumi A, et al. Anti-PL-7 antisynthetase syndrome with eosinophilic pleural effusion. *Intern Med* 2018; 57(15): 2227–2232.
68. Sharma K, Orbai AM, Desai D, et al. Brief report: antisynthetase syndrome-associated myocarditis. *J Card Fail* 2014; 20(12): 939–945.
69. Gorospe L, de la Puente-Bujidos C, Garcia-Gomez-Muriel I, et al. Asymmetric calcinosis in an adult patient with antisynthetase syndrome. *J Clin Rheumatol* 2019; 25: 36–37.
70. Aguila LA, Lopes MR, Pretti FZ, et al. Clinical and laboratory features of overlap syndromes of idiopathic inflammatory myopathies associated with systemic lupus erythematosus, systemic sclerosis, or rheumatoid arthritis. *Clin Rheumatol* 2014; 33: 1093–1098.
71. Garton MJ and Isenberg DA. Clinical features of lupus myositis versus idiopathic myositis: a review of 30 cases. *Br J Rheumatol* 1997; 36(10): 1067–1074.
72. Ghirardello A, Zampieri S, Iaccarino L, et al. Myositis specific and myositis associated autoantibodies in idiopathic inflammatory myopathies: a serologic study of 46 patients. *Reumatismo* 2005; 57(1): 22–28.
73. Hausmanowa-Petrusewicz I, Kowalska-Oledzka E, Miller FW, et al. Clinical, serologic, and immunogenetic features in Polish patients with idiopathic inflammatory myopathies. *Arthritis Rheum* 1997; 40(7): 1257–1266.
74. Hochberg MC, Feldman D and Stevens MB. Adult onset polymyositis/dermatomyositis: an analysis of clinical and laboratory features and survival in 76 patients with a review of the literature. *Semin Arthritis Rheum* 1986; 15(3): 168–178.
75. Ramesha KN, Kuruvilla A, Sarma PS, et al. Clinical, electrophysiologic, and histopathologic profile, and outcome in idiopathic inflammatory myositis: an analysis of 68 cases. *Ann Indian Acad Neurol* 2010; 13(4): 250–256.
76. Vancsa A, Gergely L, Ponyi A, et al. Myositis-specific and myositis-associated antibodies in overlap myositis in comparison to primary dermatomyositis: relevance for clinical classification: retrospective study of 169 patients. *Joint Bone Spine* 2010; 77(2): 125–130.
77. Troyanov Y, Targoff IN, Tremblay JL, et al. Novel classification of idiopathic inflammatory myopathies based on overlap syndrome features and autoantibodies: analysis of 100 French Canadian patients. *Medicine (Baltimore)* 2005; 84(4): 231–249.
78. Bhansing KJ, Lammens M, Knaapen HK, et al. Scleroderma-polymyositis overlap syndrome versus idiopathic polymyositis and systemic sclerosis: a descriptive study on clinical features and myopathology. *Arthritis Res Ther* 2014; 16(3): R111.
79. Pakozdi A, Nihtyanova S, Moizadeh P, et al. Clinical and serological hallmarks of systemic sclerosis overlap syndromes. *J Rheumatol* 2011; 38(11): 2406–2409.
80. Sato S, Hirakata M, Kuwana M, et al. Clinical characteristics of Japanese patients with anti-PL-7 (anti-threonyl-tRNA synthetase) autoantibodies. *Clin Exp Rheumatol* 2005; 23(5): 609–615.
81. Hermans MAW, Miedema JR, Verdijk RM, et al. Scleroderma-like renal crisis in a patient with anti-threonyl-tRNA synthetase-associated antisynthetase syndrome. *Rheumatology (Oxford)* 2018; 57(4): 763–765.
82. Suzuki S, Uruha A, Suzuki N, et al. Integrated diagnosis project for inflammatory myopathies: an association between autoantibodies and muscle pathology. *Autoimmun Rev* 2017; 16(7): 693–700.
83. Noguchi E, Uruha A, Suzuki S, et al. Skeletal muscle involvement in antisynthetase syndrome. *JAMA Neurol* 2017; 74(8): 992–999.
84. Aouizerate J, De Antonio M, Bassez G, et al. Myofiber HLA-DR expression is a distinctive biomarker for antisynthetase-associated myopathy. *Acta Neuropathol Commun* 2014; 2: 154.
85. Mozaffar T and Pestronk A. Myopathy with anti-Jo-1 antibodies: pathology in perimysium and neighbouring muscle fibres. *J Neurol Neurosurg Psychiatry* 2000; 68(4): 472–478.
86. Stenzel W, Preusse C, Allenbach Y, et al. Nuclear actin aggregation is a hallmark of anti-synthetase syndrome-induced dysimmune myopathy. *Neurology* 2015; 84(13): 1346–1354.
87. Mescam-Mancini L, Allenbach Y, Hervier B, et al. Anti-Jo-1 antibody-positive patients show a characteristic

- necrotizing perifascicular myositis. *Brain* 2015; 138(Pt 9): 2485–2492.
88. Uruha A, Suzuki S, Suzuki N, et al. Perifascicular necrosis in anti-synthetase syndrome beyond anti-Jo-1. *Brain* 2016; 139: e50.
 89. De Bleecker JL, De Paepe B, Aronica E, et al. 205th ENMC international workshop: pathology diagnosis of idiopathic inflammatory myopathies part II 28–30 March 2014, Naarden, The Netherlands. *Neuromuscul Disord* 2015; 25(3): 268–272.
 90. Uruha A, Allenbach Y, Charuel JL, et al. Diagnostic potential of sarcoplasmic MxA expression in subsets of dermatomyositis. *Neuropathol Appl Neurobiol* 2019; 45: 513–522.
 91. Defendenti C, Atzeni F, Spina MF, et al. Clinical and laboratory aspects of Ro/SSA-52 autoantibodies. *Autoimmun Rev* 2011; 10(3): 150–154.
 92. Zampeli E, Venetsanopoulou A, Argyropoulou OD, et al. Myositis autoantibody profiles and their clinical associations in Greek patients with inflammatory myopathies. *Clin Rheumatol* 2019; 38(1): 125–132.
 93. Schulte-Pelkum J, Fritzler M and Mahler M. Latest update on the Ro/SS-A autoantibody system. *Autoimmun Rev* 2009; 8(7): 632–637.
 94. Yamasaki Y, Satoh M, Mizushima M, et al. Clinical subsets associated with different anti-aminoacyl transfer RNA synthetase antibodies and their association with coexisting anti-Ro52. *Mod Rheumatol* 2016; 26(3): 403–409.
 95. Marie I, Hatron PY, Dominique S, et al. Short-term and long-term outcome of anti-Jo1-positive patients with anti-Ro52 antibody. *Semin Arthritis Rheum* 2012; 41(6): 890–899.
 96. Rutjes SA, Vree Egberts WT, Jongen P, et al. Anti-Ro52 antibodies frequently co-occur with anti-Jo-1 antibodies in sera from patients with idiopathic inflammatory myopathy. *Clin Exp Immunol* 1997; 109(1): 32–40.
 97. Ohashi K, Sada KE, Nakai Y, et al. Cluster analysis using anti-aminoacyl-tRNA synthetases and SS-A/Ro52 antibodies in patients with polymyositis/dermatomyositis. *J Clin Rheumatol* 2019; 25: 246–251.
 98. La Corte R, Lo Mo Naco A, Locaputo A, et al. In patients with antisynthetase syndrome the occurrence of anti-Ro/SSA antibodies causes a more severe interstitial lung disease. *Autoimmunity* 2006; 39(3): 249–253.
 99. Vancsa A, Csipo I, Nemeth J, et al. Characteristics of interstitial lung disease in SS-A positive/Jo-1 positive inflammatory myopathy patients. *Rheumatol Int* 2009; 29(9): 989–994.
 100. Bauhammer J, Blank N, Max R, et al. Rituximab in the treatment of Jo1 antibody-associated antisynthetase syndrome: anti-Ro52 positivity as a marker for severity and treatment response. *J Rheumatol* 2016; 43(8): 1566–1574.
 101. Labrador-Horrillo M, Martinez MA, Selva-O'Callaghan A, et al. Anti-MDA5 antibodies in a large Mediterranean population of adults with dermatomyositis. *J Immunol Res* 2014; 2014: 290797.
 102. Espinosa A, Dardalhon V, Brauner S, et al. Loss of the lupus autoantigen Ro52/Trim21 induces tissue inflammation and systemic autoimmunity by dysregulating the IL-23-Th17 pathway. *J Exp Med* 2009; 206(8): 1661–1671.
 103. Mileti LM, Streck ME, Niewold TB, et al. Clinical characteristics of patients with anti-Jo-1 antibodies: a single center experience. *J Clin Rheumatol* 2009; 15(5): 254–255.
 104. Rozelle A, Trieu S and Chung L. Malignancy in the setting of the anti-synthetase syndrome. *J Clin Rheumatol* 2008; 14(5): 285–288.
 105. Boleto G, Perotin JM, Eschard JP, et al. Squamous cell carcinoma of the lung associated with anti-Jo1 antisynthetase syndrome: a case report and review of the literature. *Rheumatol Int* 2017; 37(7): 1203–1206.
 106. Legault D, McDermott J, Crous-Tsanaclis AM, et al. Cancer-associated myositis in the presence of anti-Jo1 autoantibodies and the antisynthetase syndrome. *J Rheumatol* 2008; 35(1): 169–171.
 107. Przybylski G, Jarzemska A, Czerniak J, et al. A case report of a patient with dermatomyositis as a prodromal sign of lung cancer. *Pol Arch Med Wewn* 2008; 118(3): 143–147.
 108. Respicio G, Shwaiki W and Abeles M. A 58-year-old man with anti-Jo-1 syndrome and renal cell carcinoma: a case report and discussion. *Conn Med* 2007; 71(3): 151–153.
 109. Van der Bijl AE, Meinders AE, van Duinen SG, et al. A nasal squamous cell carcinoma complicated by a paraneoplastic syndrome consisting of a myositis and anti-Jo-1 autoantibodies. *Eur J Intern Med* 2005; 16(5): 369–371.
 110. Vincze M, Molnar PA, Zilahi E, et al. Primary lung adenocarcinoma associated with anti-Jo-1 positive polymyositis. *Joint Bone Spine* 2011; 78(2): 209–211.
 111. Watkins J, Farzaneh-Far R, Tahir H, et al. Jo-1 syndrome with associated poorly differentiated adenocarcinoma. *Rheumatology (Oxford)* 2004; 43(3): 389–390.
 112. Buchbinder R, Forbes A, Hall S, et al. Incidence of malignant disease in biopsy-proven inflammatory myopathy. A population-based cohort study. *Ann Intern Med* 2001; 134(12): 1087–1095.
 113. Castaneda-Pomeda M, Prieto-Gonzalez S and Grau JM. Antisynthetase syndrome and malignancy: our experience. *J Clin Rheumatol* 2011; 17(8): 458.
 114. Kobayashi S, Shoda H, Seri Y, et al. Characteristics of antisynthetase syndrome patients with hyperferritinaemia. *Scand J Rheumatol* 2017; 46(2): 161–163.
 115. Pinal-Fernandez I, Casal-Dominguez M, Huapaya JA, et al. A longitudinal cohort study of the anti-synthetase syndrome: increased severity of interstitial lung disease in black patients and patients with anti-PL7 and anti-PL12 autoantibodies. *Rheumatology (Oxford)* 2017; 56(6): 999–1007.
 116. Cruellas MG, Viana Vdos S, Levy-Neto M, et al. Myositis-specific and myositis-associated autoantibody profiles and their clinical associations in a large series of patients with polymyositis and dermatomyositis. *Clinics (Sao Paulo)* 2013; 68(7): 909–914.
 117. Kalluri M, Sahn SA, Oddis CV, et al. Clinical profile of anti-PL-12 autoantibody. Cohort study and review of the literature. *Chest* 2009; 135(6): 1550–1556.
 118. Oddis CV and Aggarwal R. Treatment in myositis. *Nat Rev Rheumatol* 2018; 14: 279–289.
 119. Conway R, Low C, Coughlan RJ, et al. Methotrexate use and risk of lung disease in psoriasis, psoriatic arthritis, and inflammatory bowel disease: systematic literature review

- and meta-analysis of randomised controlled trials. *BMJ* 2015; 350: h1269.
120. Bakewell CJ and Raghu G. Polymyositis associated with severe interstitial lung disease: remission after three doses of IV immunoglobulin. *Chest* 2011; 139(2): 441–443.
 121. Suzuki Y, Hayakawa H, Miwa S, et al. Intravenous immunoglobulin therapy for refractory interstitial lung disease associated with polymyositis/dermatomyositis. *Lung* 2009; 187(3): 201–206.
 122. Marie I, Hachulla E, Cherin P, et al. Interstitial lung disease in polymyositis and dermatomyositis. *Arthritis Rheum* 2002; 47: 614–622.
 123. Douglas WW, Tazelaar HD, Hartman TE, et al. Polymyositis-dermatomyositis-associated interstitial lung disease. *Am J Respir Crit Care Med* 2001; 164: 1182–1185.
 124. Fischer A, Brown KK, Du Bois RM, et al. Mycophenolate mofetil improves lung function in connective tissue disease-associated interstitial lung disease. *J Rheumatol* 2013; 40(5): 640–646.
 125. Hisanaga J, Kotani T, Fujiki Y, et al. Successful multi-target therapy including rituximab and mycophenolate mofetil in anti-melanoma differentiation-associated gene 5 antibody-positive rapidly progressive interstitial lung disease with clinically amyopathic dermatomyositis. *Int J Rheum Dis* 2017; 20(12): 2182–2185.
 126. Mira-Avendano IC, Parambil JG, Yadav R, et al. A retrospective review of clinical features and treatment outcomes in steroid-resistant interstitial lung disease from polymyositis/dermatomyositis. *Respir Med* 2013; 107(6): 890–896.
 127. Morganroth PA, Kreider ME and Werth VP. Mycophenolate mofetil for interstitial lung disease in dermatomyositis. *Arthritis Care Res (Hoboken)* 2010; 62: 1496–1501.
 128. Tsuchiya H, Tsuno H, Inoue M, et al. Mycophenolate mofetil therapy for rapidly progressive interstitial lung disease in a patient with clinically amyopathic dermatomyositis. *Mod Rheumatol* 2014; 24(4): 694–696.
 129. Ochi S, Nanki T, Takada K, et al. Favorable outcomes with tacrolimus in two patients with refractory interstitial lung disease associated with polymyositis/dermatomyositis. *Clin Exp Rheumatol* 2005; 23(5): 707–710.
 130. Takada K, Nagasaka K and Miyasaka N. Polymyositis/dermatomyositis and interstitial lung disease: a new therapeutic approach with T-cell-specific immunosuppressants. *Autoimmunity* 2005; 38(5): 383–392.
 131. Wilkes MR, Sereika SM, Fertig N, et al. Treatment of antisynthetase-associated interstitial lung disease with tacrolimus. *Arthritis Rheum* 2005; 52(8): 2439–2446.
 132. Kotani T, Takeuchi T, Makino S, et al. Combination with corticosteroids and cyclosporin-A improves pulmonary function test results and chest HRCT findings in dermatomyositis patients with acute/subacute interstitial pneumonia. *Clin Rheumatol* 2011; 30(8): 1021–1028.
 133. Kurita T, Yasuda S, Oba K, et al. The efficacy of tacrolimus in patients with interstitial lung diseases complicated with polymyositis or dermatomyositis. *Rheumatology (Oxford)* 2015; 54: 39–44.
 134. Kameda H, Nagasawa H, Ogawa H, et al. Combination therapy with corticosteroids, cyclosporin A, and intravenous pulse cyclophosphamide for acute/subacute interstitial pneumonia in patients with dermatomyositis. *J Rheumatol* 2005; 32(9): 1719–1726.
 135. Mok CC, To CH and Szeto ML. Successful treatment of dermatomyositis-related rapidly progressive interstitial pneumonitis with sequential oral cyclophosphamide and azathioprine. *Scand J Rheumatol* 2003; 32(3): 181–183.
 136. Yamasaki Y, Yamada H, Yamasaki M, et al. Intravenous cyclophosphamide therapy for progressive interstitial pneumonia in patients with polymyositis/dermatomyositis. *Rheumatology (Oxford)* 2007; 46(1): 124–130.
 137. Schnabel A, Reuter M, Biederer J, et al. Interstitial lung disease in polymyositis and dermatomyositis: clinical course and response to treatment. *Semin Arthritis Rheum* 2003; 32(5): 273–284.
 138. Allenbach Y, Guiguet M, Rigolet A, et al. Efficacy of rituximab in refractory inflammatory myopathies associated with anti-synthetase auto-antibodies: an open-label, phase II trial. *PLoS ONE* 2015; 10(11): e0133702.
 139. Andersson H, Sem M, Lund MB, et al. Long-term experience with rituximab in anti-synthetase syndrome-related interstitial lung disease. *Rheumatology (Oxford)* 2015; 54(8): 1420–1428.
 140. Keir GJ, Maher TM, Ming D, et al. Rituximab in severe, treatment-refractory interstitial lung disease. *Respirology* 2014; 19(3): 353–359.
 141. Kurtzman DJ, Wright NA, Lin J, et al. Tofacitinib citrate for refractory cutaneous dermatomyositis: an alternative treatment. *JAMA Dermatol* 2016; 152(8): 944–945.
 142. Paik JJ and Christopher-Stine L. A case of refractory dermatomyositis responsive to tofacitinib. *Semin Arthritis Rheum* 2017; 46(4): e19.
 143. Kato M, Ikeda K, Kageyama T, et al. Successful treatment for refractory interstitial lung disease and pneumomediastinum with multidisciplinary therapy including tofacitinib in a patient with anti-MDA5 antibody-positive dermatomyositis. *J Clin Rheumatol*. Epub ahead of print 8 January 2019. DOI: 10.1097/RHU.0000000000000984.
 144. Kurasawa K, Arai S, Namiki Y, et al. Tofacitinib for refractory interstitial lung diseases in anti-melanoma differentiation-associated 5 gene antibody-positive dermatomyositis. *Rheumatology (Oxford)* 2018; 57(12): 2114–2119.
 145. Wendel S, Venhoff N, Frye BC, et al. Successful treatment of extensive calcifications and acute pulmonary involvement in dermatomyositis with the Janus—Kinase inhibitor tofacitinib—a report of two cases. *J Autoimmun* 2019; 100: 131–136.
 146. U.S. National Library of Medicine, 2017, <https://clinicaltrials.gov/ct2/show/NCT03215927>
 147. Rowen AJ and Reichel J. Dermatomyositis with lung involvement, successfully treated with azathioprine. *Respiration* 1983; 44(2): 143–146.
 148. Sarahrudi K, Estenne M, Corris P, et al. International experience with conversion from cyclosporine to tacrolimus for acute and chronic lung allograft rejection. *J Thorac Cardiovasc Surg* 2004; 127(4): 1126–1132.
 149. Go DJ, Park JK, Kang EH, et al. Survival benefit associated with early cyclosporine treatment for dermatomyositis-associated interstitial lung disease. *Rheumatol Int* 2016; 36(1): 125–131.

150. Witt LJ, Demchuk C, Curran JJ, et al. Benefit of adjunctive tacrolimus in connective tissue disease-interstitial lung disease. *Pulm Pharmacol Ther* 2016; 36: 46–52.
151. Nakashima R and Mimori T. Anti-MDA5 (melanoma differentiation-associated gene 5) antibody and dermatomyositis with rapidly progressive interstitial pneumonia. *Nihon Rinsho Meneki Gakkai Kaishi* 2013; 36(2): 71–76.
152. Dasa O, Ruzieh M and Oraibi O. Successful treatment of life-threatening interstitial lung disease secondary to antisynthetase syndrome using rituximab: a case report and review of the literature. *Am J Ther* 2016; 23(2): e639–e645.
153. Lepri G, Avouac J, Airo P, et al. Effects of rituximab in connective tissue disorders related interstitial lung disease. *Clin Exp Rheumatol* 2016; 34(5): 181–185.
154. Nalotto L, Iaccarino L, Zen M, et al. Rituximab in refractory idiopathic inflammatory myopathies and antisynthetase syndrome: personal experience and review of the literature. *Immunol Res* 2013; 56(2–3): 362–370.
155. Sem M, Molberg O, Lund MB, et al. Rituximab treatment of the anti-synthetase syndrome: a retrospective case series. *Rheumatology (Oxford)* 2009; 48(8): 968–971.
156. Sharp C, McCabe M, Dodds N, et al. Rituximab in autoimmune connective tissue disease-associated interstitial lung disease. *Rheumatology (Oxford)* 2016; 55: 1318–1324.
157. Unger L, Kampf S, Luthke K, et al. Rituximab therapy in patients with refractory dermatomyositis or polymyositis: differential effects in a real-life population. *Rheumatology (Oxford)* 2014; 53(9): 1630–1638.
158. Vandembroucke E, Grutters JC, Altenburg J, et al. Rituximab in life threatening antisynthetase syndrome. *Rheumatol Int* 2009; 29: 1499–1502.
159. Doyle TJ, Dhillon N, Madan R, et al. Rituximab in the treatment of interstitial lung disease associated with antisynthetase syndrome: a multicenter retrospective case review. *J Rheumatol* 2018; 45(6): 841–850.
160. De Souza FHC, Miozzi R, de Moraes JCB, et al. Favorable rituximab response in patients with refractory idiopathic inflammatory myopathies. *Adv Rheumatol* 2018; 58(1): 31.
161. Aggarwal R, Bandos A, Reed AM, et al. Predictors of clinical improvement in rituximab-treated refractory adult and juvenile dermatomyositis and adult polymyositis. *Arthritis Rheumatol* 2014; 66(3): 740–749.
162. Saunders P, Tsipouri V, Keir GJ, et al. Rituximab versus cyclophosphamide for the treatment of connective tissue disease-associated interstitial lung disease (RECITAL): study protocol for a randomised controlled trial. *Trials* 2017; 18(1): 275.
163. Anh-Tu Hoa S and Hudson M. Critical review of the role of intravenous immunoglobulins in idiopathic inflammatory myopathies. *Semin Arthritis Rheum* 2017; 46(4): 488–508.
164. Lundberg IE, Tjarnlund A, Bottai M, et al. 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups. *Arthritis Rheumatol* 2017; 69: 2271–2282.